



**KAMIYA BIOMEDICAL COMPANY**

# Rat Thrombopoietin (TPO) ELISA

**For the quantitative determination of rat TPO in  
serum, plasma and other biological fluids**

**Cat. No. KT-29713**

**For Research Use Only.**

**Product Information**  
**Rat Thrombopoietin (TPO) ELISA**  
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## INTENDED USE

The kit is a sandwich enzyme immunoassay for the *in vitro* quantitative measurement of rat TPO in serum, plasma and other biological fluids. For research use only.

## COMPONENTS

Reagents	Quantity
Pre-coated, ready to use 96-well strip plate	1
Calibrator	2
Calibrator Diluent	1 × 20 mL
Detection Reagent A	1 × 120 µL
Detection Reagent B	1 × 120 µL
Assay Diluent A	1 × 12 mL
Assay Diluent B	1 × 12 mL
TMB Substrate	1 × 9 mL
Stop Solution	1 × 6 mL
Wash Buffer (30X concentrate)	1 × 20 mL
Plate sealer for 96 wells	4

## MATERIALS REQUIRED BUT NOT SUPPLIED

1. Microplate reader with 450 ± 10 nm filter.
2. Precision single or multi-channel pipettes and disposable tips.
3. Eppendorf Tubes for diluting samples.
4. Deionized or distilled water.
5. Absorbent paper for blotting the microtiter plate.
6. Container for Wash Solution

## STORAGE

All the reagents should be kept according to the labels on vials. The **Calibrator**, **Detection Reagent A**, **Detection Reagent B** and the **96-well strip plate** should be stored at -20°C upon receipt while the others should be at 4°C. The unused strips should be kept in a sealed bag with the desiccant provided to minimize exposure to damp air. Opened test kits will remain stable for 1 month provided they are stored as described above.

## PRINCIPLE

The microtiter plate provided in this kit has been pre-coated with an antibody specific to TPO. Calibrators or samples are then added to the appropriate microtiter plate wells with a biotin-conjugated antibody preparation specific for TPO. Next, Avidin conjugated to Horseradish Peroxidase (HRP) is added to each

microplate well and incubated. Then the TMB substrate solution is added to each well. Only those wells that contain TPO, biotin-conjugated antibody and enzyme-conjugated Avidin will exhibit a change in color. The enzyme-substrate reaction is terminated by the addition of a sulfuric acid solution and the color change is measured spectrophotometrically at a wavelength of 450 nm ± 10 nm. The concentration of TPO in the samples is then determined by comparing the O.D. of the samples to the calibration curve.

## SAMPLE COLLECTION AND STORAGE

### Serum

Use a serum separator tube and allow samples to clot for two hours at room temperature or overnight at 4°C before centrifugation for 20 minutes at approximately 1,000 x g. Assay freshly prepared serum immediately or store samples in aliquot at -20°C or -80°C for later use. Avoid repeated freeze/thaw cycles.

### Plasma

Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 15 minutes at 1,000 x g at 4°C within 30 minutes of collection. Remove plasma and assay immediately or store samples in aliquot at -20°C or -80°C for later use. Avoid repeated freeze/thaw cycles.

### Other biological fluids

Centrifuge samples for 20 minutes at 1,000 x g. Remove particulates and assay immediately or store samples in aliquot at -20°C or -80°C for later use. Avoid repeated freeze/thaw cycles.

### Note:

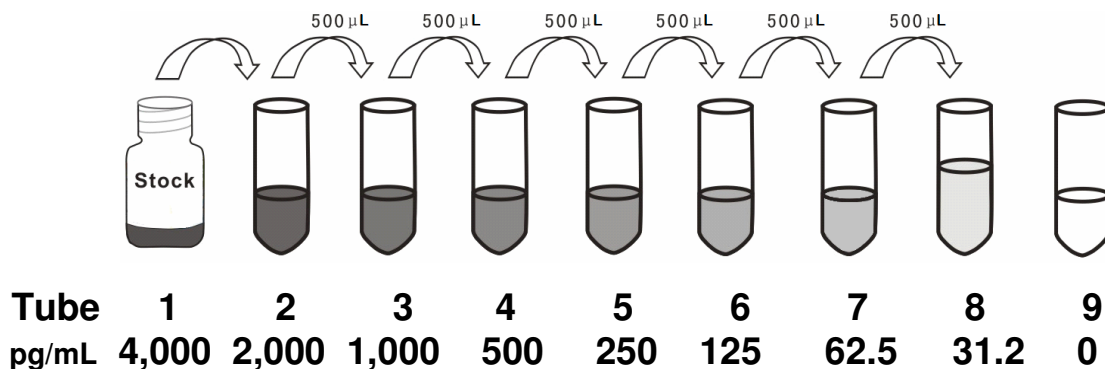
1. Samples to be used within 5 days may be stored at 4°C, otherwise samples must be stored at -20°C (≤1 month) or -80°C (≤2 months) to avoid loss of bioactivity and contamination.
2. When performing the assay, bring samples to room temperature.
3. Sample hemolysis will influence the result, so hemolytic specimen should not be detected.
4. It is highly recommended to use serum instead of plasma for the detection based on quantity of our in-house data.

## REAGENT PREPARATION

Bring all kit components and samples to room temperature (18-25°C) before use.

### Calibrator

Reconstitute the **Calibrator** with 1.0 mL of **Calibrator Diluent**, kept for 10 minutes at room temperature, shake gently (not to foam). The concentration of the calibrator in the stock solution is 4,000 pg/mL. Please firstly dilute the stock solution to 2,000 pg/mL and the diluted calibrator serves as the highest calibrator (2,000 pg/mL). Then prepare 7 tubes containing 0.5 mL Calibrator Diluent and use the diluted calibrator to produce a double dilution series according to the picture shown below. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted calibrator such as 2,000 pg/mL, 1,000 pg/mL, 500 pg/mL, 250 pg/mL, 125 pg/mL, 62.5 pg/mL, 31.2 pg/mL, and the last EP tubes with **Calibrator Diluent** is the blank as 0 pg/mL.



**Detection Reagent A and B**

Briefly spin or centrifuge the stock Detection Reagent A and Detection Reagent B before use. Dilute to the working concentration with **Assay Diluent A or B**, respectively (1:100).

**Wash Solution**

Dilute 20 mL of Wash Solution Concentrate (30X) with 580 mL of deionized or distilled water to prepare 600 mL of Wash Solution (1X).

**TMB substrate**

Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.

**Note:**

1. Prepare calibrator within 15 minutes before assay. Please do not dissolve the reagents at 37°C directly.
2. Making serial dilution in the wells directly is not permitted.
3. Please carefully reconstitute Calibrators or working Detection Reagent A and B according to the instruction, and avoid foaming and mix gently until the crystals have completely dissolved. To minimize imprecision caused by pipetting, use small volumes and ensure that pipettors are calibrated. It is recommended to suck more than 10  $\mu$ L for once pipetting.
4. The reconstituted Calibrators, Detection Reagent A and Detection Reagent B can be **used only once**.
5. If crystals have formed in the Wash Solution concentrate (30X), warm to room temperature and mix gently until the crystals have completely dissolved.
6. Contaminated water or container for reagent preparation will influence the detection result.

**SAMPLE PREPARATION**

1. Kamiya Biomedical Company is only responsible for the kit itself, but not for the samples consumed during the assay. The user should calculate the possible amount of the samples used in the whole test. Please reserve sufficient samples in advance.
2. Please predict the concentration before assaying. If values for these are not within the range of the calibration curve, users must determine the optimal sample dilutions for their particular experiments. Sample should be diluted by 0.01 mol/L PBS (PH=7.0-7.2).
3. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
4. Tissue or cell extraction samples prepared by chemical lysis buffer may cause unexpected ELISA results due to the impacts of certain chemicals.
5. Owing to the possibility of mismatching between antigen from other resource and antibody used in our kits (e.g., antibody targets conformational epitope rather than linear epitope), some native or recombinant proteins from other manufacturers may not be recognized by our products.
6. Influenced by the factors including cell viability, cell number and also sampling time, samples from cell culture supernatant may not be detected by the kit.
7. Fresh samples without long time storage is recommended for the test. Otherwise, protein degradation and denaturalization may occur in those samples and finally lead to wrong results.

**ASSAY PROCEDURE**

1. Determine wells for diluted calibrator, blank and sample. Prepare 7 wells for calibrator, 1 well for blank. Add 100  $\mu$ L each of dilutions of calibrator (read Reagent Preparation), blank and samples into the appropriate wells. Cover with the Plate sealer. Incubate for 1 hour at 37°C.

2. Remove the liquid from each well, don't wash.
3. Add 100  $\mu$ L of **Detection Reagent A** working solution to each well. Incubate for 1 hour at 37°C after covering it with the Plate sealer.
4. Aspirate the solution and wash with 350  $\mu$ L of 1X Wash Solution to each well using a squirt bottle, multi-channel pipette, manifold dispenser or autowasher, and let sit for 1~2 minutes. Remove the remaining liquid from all wells completely by snapping the plate onto absorbent paper. Repeat 3 times. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against absorbent paper.
5. Add 100  $\mu$ L of **Detection Reagent B** working solution to each well. Incubate for 30 minutes at 37°C after covering it with the Plate sealer.
6. Repeat the aspiration/wash process five times as in step 4.
7. Add 90  $\mu$ L of **Substrate Solution** to each well. Cover with a new Plate sealer. Incubate for 10 - 20 minutes at 37°C (Don't exceed 30 minutes). Protect from light. The solution will turn blue after the addition of Substrate Solution.
8. Add 50  $\mu$ L of **Stop Solution** to each well. The liquid will turn yellow after the addition of Stop solution. Mix the liquid by gently tapping the side of the plate. If color change does not appear uniform, gently tap the plate to ensure thorough mixing.
9. Remove any drops of solution or fingerprints on the bottom of the plate and confirm there are no bubbles on the surface of the liquid. Then, run the microplate reader and conduct measurements at 450 nm immediately.

**Note:**

1. **Assay preparation:** Keep appropriate numbers of strips for 1 experiment and remove extra strips from microtiter plate. Unused strips should be resealed and stored at -20°C.
2. **Sample or reagent additions:** Use freshly prepared Calibrators. Carefully add samples to wells and mix gently to avoid foaming. Do not touch the well walls if possible. For each step in the procedure, total dispensing time for addition of reagents or samples to the assay plate should not exceed 10 minutes. This will ensure equal elapsed time for each pipetting step, without interruption. Duplication of all calibrators and samples, although not required, is recommended. To avoid cross-contamination, change pipette tips between additions of each calibrator level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
3. **Incubation:** To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary. Do not allow wells to sit uncovered for extended periods between incubation steps. After reagents have been added to the well strips, DO NOT let the strips DRY at any time during the assay. Incubation time and temperature must be observed.
4. **Washing:** The wash procedure is critical. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Solution by aspirating or decanting and remove any drops of solution or fingerprints on the bottom of the plate. Insufficient washing will result in poor precision and falsely elevated absorbance reading.
5. **Controlling of reaction time:** Observe the change of color after adding **TMB Substrate** (e.g. observation once every 10 minutes), if the color is too dark, add **Stop Solution** in advance to avoid excessively strong reaction which will result in inaccurate absorbance reading.
6. **TMB Substrate** is easily contaminated. Protect from light.
7. The environment humidity which is less than 60% might have some effects on the final performance, therefore, a humidifier is recommended to be used at that condition.

**CALCULATION OF RESULTS**

Average the duplicate readings for each calibrator, control, and samples and subtract the average zero calibrator optical density. Create a calibration curve by plotting the mean OD and concentration for each

calibrator and draw a best fit curve through the points on the graph, or create a calibration curve on log-log graph paper, with TPO concentration on the y-axis and absorbance on the x-axis. Using some plot software is also recommended. If samples have been diluted, the concentration read from the calibration curve must be multiplied by the dilution factor.

## **PERFORMANCE**

### **Detection Range**

31.2 - 2,000 pg/mL.

The calibration curve concentrations used for the ELISA's were 2,000 pg/mL, 1,000 pg/mL, 500 pg/mL, 250 pg/mL, 125 pg/mL, 62.5 pg/mL, 31.2 pg/mL.

### **Sensitivity**

The minimum detectable dose of TPO is typically less than 11.9 pg/mL. The sensitivity of this assay, or Lower Limit of Detection (LLD) was defined as the lowest protein concentration that could be differentiated from zero. It was determined by the mean O.D. value of 20 replicates of the zero calibrator plus two standard deviations.

### **Specificity**

This assay has high sensitivity and excellent specificity for detection of TPO. No significant cross-reactivity or interference between TPO and analogues was observed.

Note: Limited by current skills and knowledge, it is impossible for us to complete the cross-reactivity detection between TPO and all the analogues, therefore, cross reaction may still exist.

## **ASSAY PROCEDURE SUMMARY**

1. Prepare all reagents, samples and calibrators;
2. Add 100  $\mu$ L calibrator or sample to each well. Incubate 1 hour at 37°C;
3. Aspirate and add 100  $\mu$ L prepared Detection Reagent A. Incubate 1 hour at 37°C;
4. Aspirate and wash 3 times;
5. Add 100  $\mu$ L prepared Detection Reagent B. Incubate 30 minutes at 37°C;
6. Aspirate and wash 5 times;
7. Add 90  $\mu$ L Substrate Solution. Incubate 10-20 minutes at 37°C;
8. Add 50  $\mu$ L Stop Solution. Read at 450 nm immediately.

## **IMPORTANT NOTES**

1. The final experimental results will be closely related to validity of the products, so the kit should be used prior to the expiration date. And please store the kits exactly according to the instruction.
2. Kits from different batches may be a little different in detection range, sensitivity and color developing time. Please perform the experiment exactly according to the instruction attached in kit while electronic ones from our website is only for reference.
3. Do not mix or substitute reagents from one kit lot to another. Use only the reagents supplied by manufacturer.
4. Protect all reagents from strong light during storage and incubation. All the bottle caps of reagents should be covered tightly to prevent the evaporation and contamination of microorganism. TMB Substrate should remain colorless till it is reacted with the enzyme which binds to the microplate.
5. There may be some foggy substance in the wells when the plate is opened at the first time. It will not have any effect on the final assay results. Do not remove microplate from the storage bag until needed.
6. Wrong operations during the reagents preparation and loading, as well as incorrect parameter setting for the plate reader may lead to incorrect results. A microplate reader with a bandwidth of 10 nm or less and an optical density range of 0-3 O.D. at 450  $\pm$  10nm wavelength is acceptable for use in absorbance measurement. Please read the instruction carefully and adjust the instrument prior to the experiment.
7. Variation in sample preparation and each step of experimental operation may cause different results. In order to get better reproducible results, the operation of each step in the assay should be controlled.

8. Each kit has been strictly passed Q.C test. However, results from end users might be inconsistent with our in-house data due to some unexpected transportation conditions or different lab equipments. Intra-assay variance among kits from different batches might arise from above factors, too.
9. Kits from different manufacturers with the same item might produce different results, since we haven't compared our products with other manufacturers.
10. The calibrator of the kit and immunogen used for antibody preparation are commonly recombinant proteins, as different fragments, expression systems, purification methods might be used in recombinant protein preparation, we can not guarantee the kit could detect recombinant protein from other companies. So, it is not recommended to use the kit for the detection of recombinant protein.
11. Please predict the concentration of target molecules in samples, or arrange a preliminary experiment, it is a good way to solve specific problem, e.g. the concentration of samples are beyond the detection range of the kit.
12. The kit might not be suitable for detection of samples from some special experiment, for instance, knock-out experiments, due to their uncertainty of effectiveness.
13. The Stop Solution suggested for use with this kit is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

**FOR RESEARCH USE ONLY.**

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