

KAMIYA BIOMEDICAL COMPANY

Chicken IgG ELISA

For the quantitative determination of IgG in chicken serum, plasma and egg-yolk extracts

Cat. No. KT-619

For Research Use Only.

PRODUCT INFORMATION

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PRODUCT

The **K-ASSAY®** Chicken IgG ELISA is an enzyme immunoassay for the quantitative determination of IgG in chicken serum, plasma and egg-yolk extracts. For research use only.

INTRODUCTION

The chicken IgG ELISA kit is intended for measurement of IgG in serum, plasma and egg-yolk extracts. Goat anti-chicken IgG (H+L) antibodies are used for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated goat anti-chicken IgG (Fc specific) antibodies are used for detection. Cross-reactivity with chicken IgM is ≤1%. IgM does not interfere with quantitative measurement of IgG in normal serum or plasma. Cross-reactivity with chicken IgA has not been investigated.

Normal chicken IgG levels are in the range of 3-6 mg/mL in serum or plasma and 1-2 mg/mL in egg yolk.

PRINCIPLE

Test samples are diluted and incubated in the microtiter wells for 45 minutes alongside prepared chicken IgG calibrators. The microtiter wells are subsequently washed and HRP conjugate is added and incubated for 45 minutes. IgG molecules are thus sandwiched between the immobilization and detection antibodies. The wells are then washed to remove unbound HRP-labeled antibodies and TMB Reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow, and optical density is measured spectrophotometrically at 450 nm. The concentration of IgG is proportional to the optical density of the test sample and is derived from a calibration curve.

COMPONENTS

- Anti chicken IgG coated 96-well plate (12 strips of 8 wells)
- Reference calibrator (lyophilized)
- 10X Diluent, 25 mL
- HRP Conjugate Reagent, 11 mL
- 20X Wash Solution, 50 mL
- TMB Reagent (One-step), 11 mL
- Stop Solution (1N HCl), 11 mL

MATERIALS REQUIRED BUT NOT PROVIDED

- Precision pipettes and tips
- Distilled or de-ionized water
- Vortex mixer
- Absorbent paper or paper towels
- Graph paper (PC graphing software is optional)
- Polypropylene or glass tubes
- Plate reader with an optical density range of 0-4 at 450 nm.
- Micro-Plate incubator/shaker mixing speed of ~150 rpm
- Plate washer

DILUENT PREPARATION

The diluent is provided as a 10X stock. Prior to use estimate the final volume of diluent required for your assay and dilute one (1) volume of the 10x stock with nine (9) volumes of distilled or de-ionized water.

WASH SOLUTION PREPARATION

The wash solution is provided as a 20X stock. Prior to use dilute the contents of the bottle (50 mL) with 950 mL of distilled or de-ionized water.

CALIBRATOR PREPARATION

1. The IgG calibrator is provided as a lyophilized stock. Reconstitute with 1.0 mL of distilled or de-ionized water (***the reconstituted calibrator is stable at 4°C for one week but should be aliquoted and frozen at -20°C after reconstitution if future use is intended***).
2. Label 8 polypropylene or glass tubes as 100, 50, 25, 12.5, 6.25, 3.125, 1.563 and 0 ng/mL.
3. Into the tube labeled 100 ng/mL, pipette 415.7 µL of diluent. Then add 84.3 µL of IgG calibrator and mix gently. This provides the 100 ng/mL calibrator.
4. Dispense 250 µL of diluent into the tubes labeled 50, 25, 12.5, 6.25, 3.125, 1.563 and 0 ng/mL.
5. Prepare a 50 ng/mL calibrator by diluting and mixing 250 µL of the 100 ng/mL calibrator with 250 µL of diluent in the tube labeled 50 ng/mL.
6. Similarly prepare the 25, 12.5, 6.25, 3.125 and 1.563 ng/mL calibrators by serial dilution.

SAMPLE PREPARATION

General Note: IgG is typically present in chicken serum at concentrations of ~5-10 mg/mL. In order to obtain values within range of the calibration curve, we suggest that samples initially be diluted 400,000 fold using the following procedure for each sample to be tested:

1. Dispense 999 µL and 798 µL of 1X diluent into separate tubes.
2. Pipette and mix 1 µL of the serum/plasma sample into the tube containing 999 µL of diluent. This provides a 1,000 fold diluted sample.
3. Mix 2 µL of the 1,000 fold diluted sample with the 798 µL of diluent in the second tube. This provides a 400,000 fold dilution of the sample.
4. Repeat this procedure for each sample to be tested.

ASSAY PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 µL of calibrators and diluted samples into the wells (we recommend that samples be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 µL/well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 µL of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
8. Wash as detailed in 4 to 5 above.
9. Dispense 100 µL of TMB Reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 20 minutes.
11. Stop the reaction by adding 100 µL of Stop Solution to each well.
12. Gently mix. *It is important to make sure that all the blue color changes to yellow.*
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes.

CALCULATION OF RESULTS

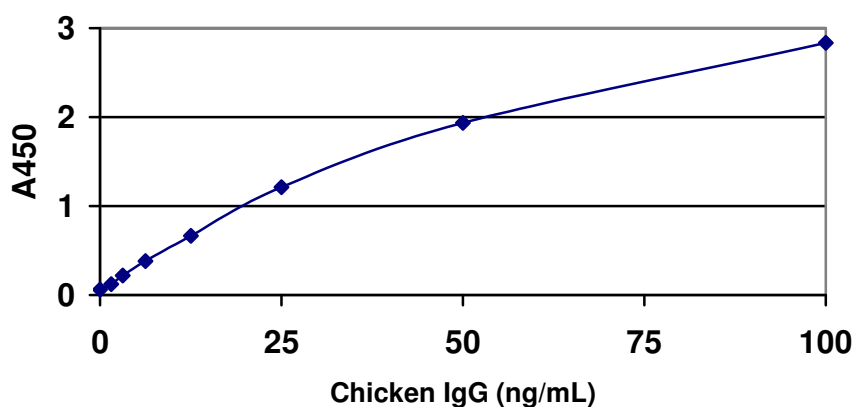
1. Calculate the average absorbance values (A_{450}) for each set of reference calibrators and samples.
2. Construct a calibration curve by plotting the mean absorbance obtained from each reference calibrator against its concentration in ng/mL on linear graph paper, with absorbance values on the vertical or Y-axis and concentrations on the horizontal or X-axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of IgG in ng/mL from the calibration curve.
4. Multiply the derived concentrations by the dilution factor to determine the actual concentration of IgG in the sample.
5. PC graphing software may be used for the above steps.
6. If the OD_{450} values of the sample fall outside the calibration curve, samples should be diluted appropriately and re-tested.

TYPICAL CALIBRATION CURVE

A typical calibration curve with optical density readings at 450 nm on the Y axis against IgG concentrations on the X axis is shown below. This curve is for the purpose of illustration only and should not be used to calculate unknowns. Each user should obtain his or her data and calibration curve in each experiment.

IgG (ng/mL)	A ₄₅₀
100	2.836
50	1.935
25	1.212
12.5	0.665
6.25	0.382
3.125	0.220
1.563	0.122
0	0.060

Typical Chicken IgG Calibration Curve



STORAGE

The test kit will remain stable until the expiration date provided that the components are stored at 4 °C. The microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air.

GENERAL INSTRUCTIONS

1. Please read and understand the instructions thoroughly before using the kit.
2. All reagents should be allowed to reach room temperature (18-25 °C) before use.
3. Optimum results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.

LIMITATIONS OF THE PROCEDURE

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

FOR RESEARCH USE ONLY

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